

# Liferiver

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## Novel Coronavirus (2019-nCoV) Real Time RT-PCR Kit User Manual

REF RR-0478-02

For use with ABI Prism®7000/7300/7500/7900/Step One Plus; iCycler iQ™4/iQ™5; Smart Cycler II; Bio-Rad CFX 96; Rotor Gene™ 6000; Mx3000P/3005P; MJ-Option2/Chromo4; LightCycler®480 Instrument



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### 1. Intended Use

Novel Coronavirus (2019-nCoV) Real Time RT-PCR Kit is used for the qualitative detection of a novel coronavirus, which was identified in 2019 at Wuhan City, Hubei Province, China, in upper respiratory tract specimens (nasopharyngeal extracts, deep cough sputum, etc.) and lower respiratory tract specimens (alveoli irrigation fluid, etc.) by real time PCR systems.

### 2. Principle of Real-Time PCR

The principle of the real-time detection is based on the fluorogenic 5' nuclease assay. During the PCR reaction, the DNA polymerase cleaves the probe at the 5' end and separates the reporter dye from the quencher dye only when the probe hybridizes to the target DNA. This cleavage results in the fluorescent signal generated by the cleaved reporter dye, which is monitored real-time by the PCR detection system. The PCR cycle at which an increase in the fluorescence signal is detected initially (Ct) is proportional to the amount of the specific PCR product. Monitoring the fluorescence intensities in real time allows the detection of the accumulating product without having to re-open the reaction tube after the amplification.

### 3. Product Description

On January 11, 2020, Chinese health authorities preliminarily identified more than 40 human infections with a novel coronavirus in an outbreak of pneumonia under investigation in Wuhan City, Hubei Province, China. The Chinese authorities identified a new type of coronavirus (novel coronavirus, named as 2019-nCoV), which was isolated on 7 January 2020.

Coronaviruses are a large family of viruses, some causing illness in human and others circulating among animals such as camels, cats and bats. 2019-nCoV is a novel coronavirus. The primer and probe design for this kit is based on the newly released strain (2019-nCoV) (GeneBank accession: MN908947) and covers 6 2019-nCoV strains sequences (EPI\_ISL\_402119, EPI\_ISL\_402120, EPI\_ISL\_402121, EPI\_ISL\_402122, EPI\_ISL\_402123, EPI\_ISL\_402124 included).

The kit contains a specific ready-to-use system for the detection of Novel Coronavirus (2019-nCoV) by Reverse transcription Polymerase Chain Reaction (RT-PCR) in the real-time PCR system. The master contains a Super Mix for the specific amplification of virus RNA. The reaction is done in one step real time RT-PCR. The first step is a reverse transcription (RT), during which the virus RNA is transcribed into cDNA. Afterwards, a thermostable DNA polymerase is used to amplify the specific gene fragments by means of polymerase chain reaction (PCR). Fluorescence is emitted and measured by the real time systems' optical unit during PCR. The detection of amplified virus DNA fragment is performed in fluorimeter channel FAM and channel HEX/VIC/JOE with the fluorescent quencher BHQ1. In addition, the kit contains a system to identify possible PCR inhibition by measuring the Cal Red 610/ROX/TEXAS RED fluorescence of the internal control (IC).

### 4. Kit Contents

Ref.	Type of reagent	Presentation	25rxns
1	Novel CoV (2019-nCoV) Super Mix	1 vial, 480µl	
2	RT-PCR Enzyme Mix	1 vial, 28µl	
3	Novel CoV (2019-nCoV) Internal Control	1 vial, 30µl	
4	Novel CoV (2019-nCoV) Negative Control	1 vial, 400µl	
5	Novel CoV (2019-nCoV) Positive Control	1 vial, 400µl	

### Analysis sensitivity: 1 X 10<sup>3</sup> copies/ml;

Note: Analysis sensitivity depends on the sample volume, elution volume, nucleic acid extraction method and other factors. If you use the RNA extraction kits recommended, the analysis sensitivity is the same as it declares. However, when the sample volume is dozens or even hundreds of times greater than elution volume by some concentrating method, it can be much higher.

### 5. Storage

- All reagents should be stored at -20 °C. Storage at +4 °C is not recommended.
- All reagents can be used until the expiration date indicated on the kit label.
- Repeated thawing and freezing (> 3x) should be avoided, as this may reduce the sensitivity of the assay.
- Cool all reagents during the working steps.
- Super Mix should be stored in the dark.

### 6. Additionally Required Materials and Devices

- Biological cabinet
- Vortex mixer
- Cryo-container
- Sterile filter tips for micro pipets
- Disposable gloves, powderless
- Refrigerator and Freezer
- Desktop microcentrifuge for "eppendorf" type tubes (RCF max. 16,000 x g)
- Real time PCR system
- Real time PCR reaction tubes/plates
- Pipets (0.5µl – 1000µl)
- Sterile microtubes
- Biohazard waste container
- Tube racks

### 7. Warnings and Precaution

- Carefully read this instruction before starting the procedure.
- This assay needs to be carried out by skilled personnel.
- Clinical samples should be regarded as potentially infectious materials and should be prepared in a laminar flow hood.
- This assay needs to be run according to Good Laboratory Practice.
- Do not use the kit after its expiration date.
- Avoid repeated thawing and freezing of the reagents, this may reduce the sensitivity of the test.
- Once the reagents have been thawed, vortex and centrifuge briefly the tubes before use.
- Prepare quickly the Reaction mix on ice or in the cooling block.
- Set up two separate working areas: 1) Isolation of the RNA/ DNA and 2) Amplification/

detection of amplification products.

- Pipets, vials and other working materials should not circulate among working units.
- Use always sterile pipette tips with filters.
- Wear separate coats and gloves in each area.
- Do not pipette by mouth. Do not eat, drink, smoke in laboratory.
- Avoid aerosols

### 8. Sample Collection, Storage and transport

- Collected samples in sterile tubes;
- Specimens can be extracted immediately or frozen at -20 °C to -80 °C.
- Transportation of clinical specimens must comply with local regulations for the transport of etiologic agents

### 9. Procedure

#### 9.1 RNA-Extraction

Different brand RNA extraction kits are available. You may use your own extraction systems or the commercial kits based on the yield. For the RNA extraction, please follow the manufacturer's instructions. The recommended extraction kits are as follows:

Nucleic Acid Isolation Kit	Cat. Number	Manufacturer
RNA Isolation Kit	ME-0010/ME-0012 /ME-0044	ZJ Biotech
QIAamp Viral RNA Mini extraction Kit (50)	52904	QIAGEN

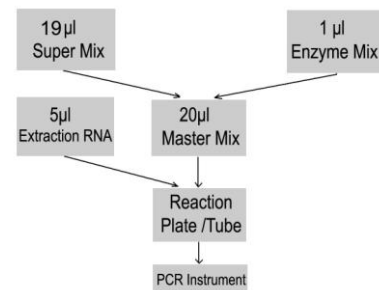
It is noted that the Novel CoV (2019-nCoV) positive control and Novel CoV (2019-nCoV) negative control in this kit should be extracted with the same protocol for specimens.

#### 9.2 Internal Control

The internal control in this kit should be added into the extraction mixture with 1µl/test to monitor the whole process. The internal control in this kit should be added into the extraction mixture with 1µl/test to monitor the whole process.

#### 9.3 RT-PCR Protocol

The Master Mix volume for each reaction should be pipetted as follows:



- The volumes of Super Mix and Enzyme Mix per reaction multiply with the number of samples, which includes the number of controls, standards, and sample prepared. Molecular Grade Water is used as the negative control. For reasons of unprecise pipetting, always add an extra virtual sample. Mix completely then spin down briefly with a centrifuge.
- Pipet 20µl Master Mix with micropipets of sterile filter tips to each of the Real time PCR reaction plate/tubes. Separately add 5µl RNA template (nucleic acid extracted from negative control, positive control and specimens) to different reaction plate/tubes. Immediately close the plates/tubes to avoid contamination.
- Spin down briefly in order to collect the Master Mix in the bottom of the reaction tubes.
- Perform the following protocol in the instrument:

45 °C for 10min	1cycle
95 °C for 15min	1cycle
95 °C for 15sec, 60 °C for 1min ( Fluorescence measured at 60 °C )	45cycles

Selection of fluorescence channels	
FAM	Target Gene
HEX/VIC/JOE	Target Gene
Cal Red 610/ROX/TEXAS RED	IC

- If you use ABI Prism® system, please choose "none" as passive reference and quencher.

**10.Threshold Setting:** Just above the maximum level of molecular grade water.

**11.Quality control:** Negative control, and positive control must be performed correctly, otherwise the sample results are invalid.

Channel	Ct value		
	FAM	HEX/VIC/JOE	Cal Red 610
Control	UNDET	UNDET	25-43
Negative Control	UNDET	UNDET	25-43
Positive Control	<35	<35	25-43

### 12. Data Analysis and Interpretation

The following sample results are possible:

	Ct value			Result Analysis
	FAM	HEX/VIC/JOE	Cal Red 610	
1#	UNDET	UNDET	25~43	Below the detection limit or negative
2#	≤43	---	---	If the Ct value of either Channel FAM or Channel HEX/VIC/JOE is ≤43, the specimen is 2019-nCoV positive.
	---	≤43	---	
3#		43~45	25~43	Re-test; If it is still 43~45, report as 1#
4#		UNDET	UNDET	PCR Inhibition; No diagnosis can be concluded.

For further questions or problems, please contact our technical support at [trade@liferiver.com.cn](mailto:trade@liferiver.com.cn)